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Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713597286

SYNTHESIS of 2'-DEOXY-2'-FLUOROGUANYL-(3',5')-GUANOSINE

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Online publication date: 12 February 2002

 $\label{thm:continuous} \textbf{To cite this Article} \ \ Maruyama, \ Tokumi \ , \ Kozai, \ Shigetada \ , \ Nakamura, \ Kazuo \ and \ Irie, \ Masachika (2002) \ 'SYNTHESIS \ of 2'-DEOXY-2'-FLUOROGUANYL-(3',5')-GUANOSINE' \ , \ Nucleosides, \ Nucleotides \ and \ Nucleic \ Acids, \ 21:11,765-774$

To link to this Article: DOI: 10.1081/NCN-120016479 URL: http://dx.doi.org/10.1081/NCN-120016479

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NUCLEOSIDES, NUCLEOTIDES & NUCLEIC ACIDS Vol. 21, Nos. 11 & 12, pp. 765–774, 2002

SYNTHESIS OF 2'-DEOXY-2'-FLUOROGUANYL-(3',5')-GUANOSINE

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ABSTRACT

The protected analogue of 2-amnio-6-chloropurine arabinoside (**3b**) was subjected to reaction with diethylaminosulfur trifluoride (DAST) and subsequently treated with NaOAc in Ac₂O/AcOH to give N^2 , $O^{3'}$, $O^{5'}$ -triacetyl-2'-deoxy-2'-fluoroguanosine (**5a**). After deacetylation of the sugar moiety and protection of 5'-OH by a 4,4'-dimethoxytrityl group, this nucleoside component was converted to 2'-deoxy-2'-fluoroguanyl-(3',5')-guanosine (**6c**, GfpG).

Ribonucleases (RNases) are very important enzymes for RNA metabolism in almost all organisms; they include those that hydrolyze only single-stranded RNA, double-stranded RNA, and RNA hybridized with DNA. The mode of hydrolysis is thought to be via a 2',3'-cyclic phosphated intermediate at the 3'-terminus of oligonucleotides, ultimately forming oligo- or mononucleotides with a terminal 3'-phosphated (transferase-type RNase). Recently, some

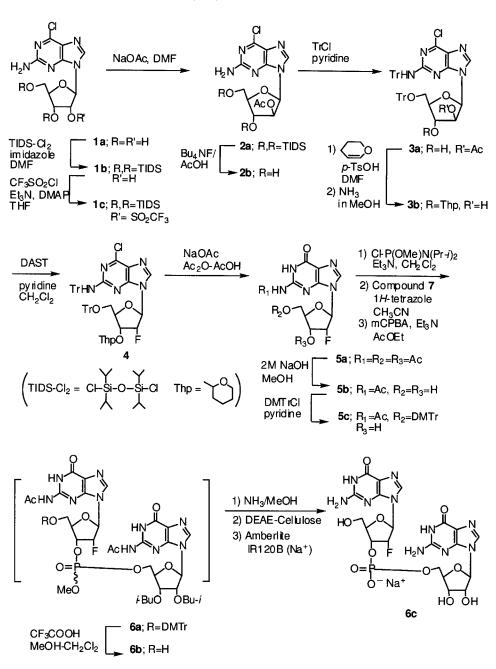
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ribonucleases showed biological activity in addition to digestion of RNAs. RNase A was demonstrated to show toxicity to tumor cells, both in vitro^[1] and in vivo.^[2,3] Also, onconase, discovered from the Northern lieopard frog,^[4] has emerged as a promising cancer chemotherapeutic agent. To elucidate the structure-function relationship of ribonucleases, X-ray crystallographic analysis of ribonucleases complexed with the analogues of dinucleoside monophosphate was achieved in many laboratories,^[5–8] An especially important analogue is 2'-deoxy-2'-fluoroguanyl-(3',5')-cytidine (GfpC),^[9] in which the 2'-hydroxyl group (2'-OH) of guanosine is replaced by a 2'-fluorine atom to prevent the transesterification step.

The synthesis of 2'-deoxy-2'-fluoronucleosides has been accomplished by several routes including nucleophilic displacement of 2'-O-trifluoro-methanesulfonylarabinosides. Recently, a method to introduce fluorine into the sugar moiety of guanine nucleosides using diethylaminosulfur trifluoride (DAST) has been developed. However, problems still remain in respect of the yields and the analogues modified at the base moiety. This background prompted us to develop a new method to introduce fluorine at the downside of the 2'-carbon of guanosine. This paper reports a method for the synthesis of 2'-deoxy-2'-fluoroguanosine in which treatment of the protected 2-amino-6-chloropurine arabinoside with DAST was involved as a key step.

An attempt at the conversion of 1a to the 2'-O-acetyl arabinoside (2) was made by application of the method of Fukukawa et al. [13] Thus, treatment of **1a** with 1,3-dichloro-1,1,3,3-tetraisopropyldisiloxane afforded the corresponding 3',5'-O-(tetraisopropyldisiloxan-1,3-diyl) derivative **1b**.^[14] Compound 1b was allowed to react with trifluoromethanesulfonyl chloride in the presence of Et₃N and 4-dimethylaminopyridine in THF to give the 2'-O-triflate 1c. An S_N2 displacement with the acetate anion at the 2'position of 1c was employed to obtain the 2'(S)(ara)-O-acetate 2a. At this stage, the silyl protecting group should be changed to a group stable to the fluoride anion. Desilylation of 2a with 2 equivalents of tetrabutylammonium fluoride in the presence of acetic acid at 0°C for 15 min afforded **2b.** [15] Since acyl protection causes neighboring group participation in the nucleophilic substitution, ethereal groups were chosen as a protecting group. [16] Reaction of 2b with trityl chloride in pyridine gave a ditrityl derivative (3a), which was successively treated with 3,4-dihydro-2*H*-pyran and ammonia in methanol to give the N^2 . O^{5'}-ditrityl-2-amino-6-chloro-[3-O-(tetrahydro-2-pyranyl)-9-β-D-arabinofuranosyl]-purine (3b). Then, 3b was treated with DAST in the presence of pyridine in CH₂Cl₂ to afford the 2'fluoride 4 in 60% yield, a key intermediate for the synthesis of basemodified analogues. No spot other than 4 was observed on TLC. When the DAST reaction was tried in the absence of pyridine, the protecting groups were removed from 3b and 4 to give a complicated result. Reaction to convert the 6-chloro atom of 4 to 6-oxo was carried out by treatment with



Scheme 1.

Figure 1. NOESY spectrum of 5b.

NaOAc in Ac₂O-AcOH. Also, two trityl groups and the tetrahydro-2-pyranyl group were removed in acidic medium, followed by acetylation with Ac₂O to afford N^2 , $O^{3'}$, $O^{5'}$ -triacetyl-2'-deoxy-2'-fluoroguanosine (**5a**). Compound **5a** was subjected to partial deacylation with 2M NaOH in MeOH at 0°C to afford **5b**. The ¹H-NMR spectrum of **5b** indicated that the 2'-fluorine caused a downfield shift of the 2'-proton and a large H2'-C-F geminal coupling (52.7 Hz), and the nuclear Overhauser effect (NOE) was observed between H2' and H3' in the two-dimensional NOE (NOESY) spectrum of **5b** (Fig. 1). Therefore, the structure of **5b** was unequivocally determined. Then, a 4,4'-dimethoxytrityl group was introduced at 5'-OH of **5b** to give a nucleoside component **5c**.

2'-Deoxy-2'-fluoroguanyl-(3',5')-guanosine (6c) was prepared according to the established phosphite-triester method. [17] Thus, 5c was treated with N,N-diisopropyl-methylphosphonamidic chloride in the presence of triethylamine in dry CH₂Cl₂. Monitoring the phosphitylating reaction by thin-layer chromatography (TLC) revealed the appearance of the 3'-O-(N,N-diisopropylamino)phosphramidites as a mixture of diastereomers. The products were converted to the monotetrazolides in situ and condensed with N^2 -acetyl-2',3'-di-O-isobutyrylguanosine (7) to form the dinucleotide phosphite triester. Mild oxidation and separation by silica gel chromatography furnished the full-protected phosphate triester 6a as a caramel in 55% overall yield from 5c. The product was deprotected successively with CF₃COOH in MeOH and ammonia in MeOH and separated by DEAE-cellulose chromatography using 0-0.12 M triethylammonium bicarbonate (TEAB) as eluent. After removal of the solution, the product was passed through a column of Amberlite IR120B (Na⁺) to give 2'-deoxy-2'-fluoroguanyl-(3',5')-guanosine (6c, Na⁺ salt) as a film in 34% overall yield from 5c. The structure of the product was determined by the presence of two sets of sugar-protons in ¹H NMR and FAB mass spectroscopy (FAB-MS).

We are looking forward to obtaining the crystal structure of ribonuclease complexed with 2'-deoxy-2'-fluoroguanyl-(3',5')-guanosine.

EXPERIMENTAL

Melting points (mp) were determined using a Yanagimoto micromelting point apparatus (hot stage type) and are uncorrected. UV spectra were recorded with a Shimadzu UV-190 digital spectrometer. Low-resolution mass spectra were obtained on a Shimadzu-LKB 9000B mass spectrometer in the direct-inlet mode. High-resolution mass spectra were obtained on a JMS AX-500 spectrometer in the direct-inlet mode. ¹H-NMR spectra were recorded on either Varian UNITY 200 (200 MHz) or Varian UNITY 600 (600 MHz) in CDCl₃ (or dimethyl sulfoxide (DMSO)-*d*₆) with tetramethylsilane as an internal standard. Merck Ark 5554 plates precoated with silica gel 60 containing fluorescent indicator F₂₅₄ were used for thin-layer chromatography and silica gel 60 (Merck 7734, 60–200 mesh) was employed for column chromatography.

9-(2-O-Acetyl-3,5-O-tetraisopropyldisiloxan-1,3-diyl)-β-D-arabinofuranosyl)-2-amino-6-chloropurine (2a). 2-Amino-6-chloro-9-(3,5-O-tetraisopropyldisiloxan-1,3-diyl-β-D-ribofuranosyl)purine (**1b**) was prepared from 2-amino-6-chloro-purine riboside (1a) according to the published method. [14] To an ice-cooled solution of **1b** (19.4 g, 35.7 mmol), triethylamine (8.8 mL), and 4-dimethylaminopyridine (7.1 g, 58 mmol) in CH₂Cl₂ (300 mL) was added trifluoromethanesulfonyl chloride (7.1 mL, 66.7 mmol) and the solution was kept at room temperature for 10 min, then water (200 mL) was added. The organic layer was dried over MgSO₄, and concentrated to a small volume. The residual solution was chromatographed over a column of silica gel G using 25% AcOEt in hexane. Evaporation of the fraction gave 1c as a caramel, which was immediately dissolved in DMF (150 mL) and sodium acetate (12 g, 146 mmol) was added. The solution was stirred at 50°C for 10 h, then partitioned between benzene (500 mL) and water (500 mL). The organic layer was washed with water twice (500 mL), dried over MgSO₄, and concentrated to a small volume. The residual solution was chromatographed over a column of silica gel G using 20–33% AcOEt in hexane. Evaporation of the fraction gave 2a as a caramel (13.4 g, 64%). ¹H NMR (CDCl₃) δ 8.04 (1H, s, H8), 6.35 (1H, d, J = 6.3 Hz, H1'), 5.57 (1H, dd, J = 6.3, 8.2 Hz, H2'),5.13 (2H, br s, NH₂), 4.69 (1H, dd, J = 8.2, 8.5 Hz, H3'), 4.03–4.18 (2H, m, H5'), 3.88–3.92 (1H, m, H4'), 1.78 (3H, s, Ac), 0.97–1.18 (28H, m, (CH₃)₂-CH x 4); UV λ_{max} (MeOH) nm: 249, 310; EI-MS m/z: 585, 587[M⁺].

9-(2-O-Acetyl-β-D-arabinofuranosyl)-2-amino-6-chloropurine (2b). To a solution of 2a (13.53 g, 23.1 mmol) and AcOH (2.86 mL, 50 mmol) in dry THF (50 mL) was added 1 M tetrabutylammonium fluoride (50 mL) and the solution was stirred at room temperature for 2 h, ten concentrated to a small volume. The residual solution was chromatographed over a column of silica gel G with 0–17% EtOH in CHCl₃. Evaporation of the fraction gave a

caramel (7.36 g, 93%). ¹H NMR (DMSO- d_6) δ 8.25 (1H, s, H8), 7.02 (2H, br s, NH₂), 6.30 (1H, d, J=5.5 Hz, H1′), 5.87 (1H, br s, 3′OH), 5.24 (1H, dd, J=5.5, 5.8 Hz, H2′), 5.07 (1H, br s, 5′OH), 4.37 (1H, t, J=5.8 Hz, H3′), 3.83–3.88 (1H, m, H4′), 3.61–3.74 (2H, m, H5′), 1.79 (3H, s, Ac); UV λ_{max} (MeOH) nm: 249, 310; EI-MS m/z: 343, 345 [M⁺]. HR-MS m/z: 343.0689 (M⁺, C₁₂H₁₄CIN₅O₅ requires 343.0684).

 N^2 , $O^{5'}$ -Ditrityl-9-(2-O-acetyl- β -D-arabinofuranosyl)-2-amino-6-chloropurine (3a). To a solution of 2b (9.16 g, 26.7 mmol), 4-dimethylaminopyridine (1.22 g, 10 mmol) and triethylamine (13.3 mL, 95 mmol) in dry DMF (220 mL) was added trityl chloride (27.8 g, 100 mmol). The solution was stirred at 50°C overnight, then MeOH (50 mL) was added and stirring was continued at room temperature for 1 h. The solvents were removed under reduced pressure and the residue was partitioned between CHCl₃ (300 mL) and water (100 mL). The organic layer was washed with water (100 mL), dried over MgSO₄, and concentrated to a small volume. The residual solution was chromatographed over a column of silica gel G using 25-50% AcOEt in hexane. Evaporation of the fraction gave a caramel (15.34 g, 69%). ¹H NMR (CDCl₃) δ 7.77 (1H, s, H8), 7.18–7.45 (ca. 30H, m, Tr×2), 6.63 (1H, br s, H1'), 5.71 (1H,br s, N^2 -H), 4.55 (1H, br s, 3'OH), 4.16–4.18 (1H, m, H3'), 3.98 (1H, q, J=4.9 Hz, H4'), 3.35 (2H, d, J=4.9 Hz, H5'), 2.72 (1H, d, J = 2.7 Hz, H2'), 1.73 (3H, s, Ac); UV λ_{max} (MeOH) nm: 259, 313; FAB-MS m/z: 828 [M⁺], 850 [M⁺+Na].

 N^2 , $O^{5'}$ -Ditrityl-2-amino-6-chloro-9-[3-O-(tetrahydro-2-phranyl)-β-D-arabinofuranosyl]purine (3b). To an ice-cooled solution of 3a (8.28 g, 10 mmol) and 3,4-dihydro-2H-pyran (5 mL, 55 mmol) in dry DMF (50 mL) was added p-toluenesulfonic acid (800 mg, 4.2 mmol), and the solution was kept at 4°C overnight, then neutralized with triethylamine. The solvents were removed under reduced pressure and the residue was partitioned between benzene (300 mL) and water (300 mL). The organic layer was washed with water twice (300 mL), dried over MgSO₄, and concentrated to a small volume. The residual solution was chromatographed over a column of silica gel G using 17–25% AcOEt in hexane. Evaporation of the fraction gave a caramel, which was dissolved in CH₂Cl₂ (150 mL). To the solution was added ammonia in MeOH (100 mL), and the solution was kept at 4°C overnight. The solvents were removed under reduced pressure to give 3b as a caramel (6.99 g, 80%). UV λ_{max} (MeOH) nm: 255, 313; FAB-MS m/z: 870, 872 [M⁺], 892, 894 [M⁺+Na]. HR-MS m/z: 870.3436 (M⁺, C₅₃H₄₈ClN₅O₅ requires 870.3420).

 N^2 , $O^{5'}$ -Ditrityl-2-amino-6-chloro-9-[2-deoxy-2-fluoro-3-O-(tetrahydro-2-pyranyl)- β -D-ribofuranosyllpurine (4). To an ice-cooled solution of 3b (13.52 g, 15.5 mmol) and pyridine (6.3 mL) in dry CH₂Cl₂ (200 mL) was added diethylaminosulfur trifluoride (3.3 mL, 25 mmol), and the solution was

stirred at 50°C for 6h. The whole was added dropwise to 5% NaHCO₃ (500 mL) with stirring then the organic layer was diluted with CH₂Cl₂ (500 mL), washed water twice (500 mL), dried over MgSO₄, and concentrated to a small volume. The residual solution was chromatographed over a column of silica gel G using 15–40% AcOEt in hexane. Evaporation of the fraction gave white crystals (8.12 g, 60%). mp 201–201.5°C; UV λ_{max} (MeOH) nm: 260, 314; FAB-MS m/z: 872, 874 [M⁺], 894, 896 [M⁺+Na]; Anal Calcd for C₅₃H₄₇ClFN₅O₄: C, 72.97; H, 5.42; N, 8.03. Found: C, 72.78; H, 5.52; N, 7.90.

 N^2 , O^3 , O^5 -Triacetyl-2'-deoxy-2'-fluoroguanosine (5a). A solution of 4 (4.73 g, 5.40 mmol) and sodium acetate (2.4 g, 30 mmol) in mixture of acetic anhydride (50 mL) and acetic acid (50 mL) was stirred at 120°C overnight. Then the solvents were removed under reduced pressure and the residue was dissolved in EtOH (30 mL). The solution was kept at room temperature for 3 hrs and concentrated to a small volume. The residue was partitioned between CHCl₃ (300 mL) and water (100 mL). The organic layer was washed with 5% NaHCO₃ (150 mL) and water (100 mL), dried over MgSO₄, and concentrated to a small volume. The residual solution was chromatographed over a column of silica gel G using 0-10% EtOH in CHCl₃. Evaporation of the fraction and crystallization from MeOH gave white crystals (1.79 g, 80%). Mp 108–108.5°C; ¹H NMR (CDCl₃) δ 12.08 (1H, s, N¹-H), 9.72 (1H, S, N^2 -H), 7.78 (1H, s, H8), 6.04 (1H, m, H1'), 5.56–5.68 (2H, m, H2', H3'), 4.70 (1H, dd, J = 4.7, 12.0 Hz, H5'a), 4.48-4.50 (1H, m, H4'), 4.25 (1H, dd, height)J = 5.5, 12.0 Hz, H5'b), 2.32 (3H, s, Ac), 2.18 (3H, s, Ac), 2.07 (3H, s, Ac); UV λ_{max} (MeOH) nm: 259, 280; EI-MS m/z: 411 [M⁺]. HR-MS m/z: 411.1181 (M^+ , $C_{16}H_{18}FN_5O_7$ requires 411.1203).

*N*²-Acetyl-2'-deoxy-2'-fluoroguanosine (5b). To an ice-cooled solution of 5a (1.40 g, 3.4 mmol) in MeOH (80 mL) was added 1 M NaOH (15 mL), and the solution was stirred at 0°C for 10 min. Then the solution was neutralized with 1 M HCl and concentrated to a small volume. The residual solution was crystallized from water (10 mL) to give white crystals (970 mg, 87%). Mp 235.5–237°C; ¹H-NMR (DMSO- d_6) δ 12.08 (1H, s, N^1 -H), 11.74 (1H, s, N^2 -H), 8.27 (1H, s, H8), 6.11 (1H, DD, J = 2.5, 16.5 Hz, H1'), 5.71 (1H, d, J = 6.0 Hz, 3'OH), 5.30 (1H, ddd, J = 2.5, 4.1, 52.7 Hz, H2'), 5.18 (1H, t, J = 5.2 Hz, 5'OH), 4.38–4.44 (1H, m, H3'), 3.95–3.97 (1H, m, H4'), 3.75–3.78 (1H, m, H5'a), 3.59–3.63 (1H, m, H5'b), 2.19 (3H, s, Ac); UV λ_{max} (MeOH) nm: 259, 280; *Anal* Calcd for $C_{12}H_{14}FN_5O_5 \cdot 0.2H_2O$: C, 43.56; H, 4.38; N, 21.17. Found: C, 43.58; H, 3.95; N, 21.01.

 N^2 -Acetyl-5'-O-(4,4'-dimethoxytrityl)-2'-deoxy-2'-fluoroguanosine (5c). To a solution of 5b (940 mg, 2.87 mmol) in pyridine (50 mL) was added

4,4'-dimethoxytrityl chloride (2.0 g, 6 mmol) and stirred at room temperature for 3 h. Water (5 mL) was added to the solution and concentrated to a small volume. The residue was partitioned between CHCl₃ (150 mL) and water (70 mL) and the organic layer was dried over MgSO₄, then concentrated to a small volume. The residue was evaporated azeotropically with toluene and chromatographed over a column of silica gel G using 0–10% EtOH in CHCl₃. Evaporation of the fraction and crystallization from benzene gave white crystals (1.39 g, 77%). Mp 164.5–165.5°C; 1 H NMR (DMSO- d_{6}) δ 12.08 (1H, s, N^{1} -H), 11.68 (1H, s, N^{2} -H), 8.12 (1H, s, H8), 6.77–7.37 (ca.13H, m, phenyl of DMTr), 6.20 (1H, dd, J=1.4, 18.9 Hz, H1'), 5.68 (1H, d, J=7.1 Hz, 3'OH), 5.41 (1H, ddd, J=1.4, 4.4 52.5 Hz, H2'), 4.54–4.61 (1H, m, H3'), 4.11–4.13 (1H, m, H4'), 3.723 (3H, s, OCH₃), 3.719 (3H, s, OCH₃), 3.26–3.33 (2H, m, H5'), 2.18 (3H, s, Ac); UV λ_{max} (MeOH) nm: 237, 281; FAB-MS m/z: 630 [M⁺+H], 652 [M⁺+Na]. HR-MS m/z: 652.2178 (M⁺+Na,C₃₃H₃₂FN₅O₇ requires 652.2179).

2'-Deoxy-2'-fluoroguanyl-(3',5')-guanosine (6, Na⁺ salt). To a solution of 5c (232 mg, 0.368 mmol) and triethylamine (0.1 mL) in CH₂Cl₂ (6 mL) was added N,N-diisopropylmethylphosphonamidic chloride (0.11 mL, 0.57 mmol) and the solution was stirred at room temperature under Ar atmosphere for 40 min, then concentrated to a small volume. The residue was partitioned between AcOEt (12 mL) and water (2 mL) and the organic layer was washed with water (2 mL), dried over MgSO₄, and evaporated. The residue was dissolved in CH₃CN (6 mL) and 1*H*-tetrazole (64 mg, 0.91 mmol) and N^2 acetyl-2',3'-di-O-isobutyrylguanosine (343 mg, 0.74 mmol) was added successively to the solution. After stirring at room temperature overnight, the solution was evaporated and the residue was partitioned between AcOEt (15 mL) and water (1.5 mL). The organic layer was washed with water (1.5 mL), dried over MgSO₄, filtered to remove insoluble materials. To the filtrate was added 3-chloroperbenzoic acid (300 mg, 1.7 mmol) and triethylamine (0.15 mL), and the solution was stirred at room temperature for 3 h. Then the solvents were concentrated to a small volume and chromatographed over a column of silica gel G using 0–10% MeOH in AcOEt. Evaporation of the fraction gave full-protected GfpG **6a** as a caramel (237 mg, 55% from **5c**). UV λ_{max} (MeOH) nm: 254, 277; FAB-MS m/z: 1193 [M⁺+Na], 1209 [M⁺+K]. The product **6a** (212 mg, 0.178 mmol) was dissolved in a mixture of MeOH (6 mL) and CH₂Cl₂ (2 mL), then trifluoroacetic acid (0.1 mL) was added. The solution was stirred at room temperature for 3 h, and neutralized with triethylamine. The solvents was removed under reduced pressure, and the residue was chromatographed over a column of silica gel G using 0–10% EtOH in CHCl₃. Evaporation of the fraction gave the partially deprotected product as a white powder **6b** (112 mg, 72%). UV λ_{max} (MeOH) nm: 260, 277; FAB-MS m/z: 869 [M⁺+H], 891 [M⁺+Na]. A part of the product (80 mg, 92 µmol) was dissolved in MeOH (15 mL) and the solution was

saturated with ammonia at 0°C. The solution was stirred at room temperature overnight and concentrated to a small volume. The residue was chromatographed over a column of DEAD-cellulose ($2 \times 20 \,\mathrm{cm}$) using 0–0.12 M TEAB (1 L). The main fraction was evaporated under reduced pressure and the residue was evaporated azeotropically with water. The residue was dissolved in water (2 mL) and passed through the column of Amberlite IR 120 (Na⁺ form, 10 mL). Evaporation of water (50 mL) gave 6 as a filmy substance (**6c**, 51 mg, 78.2 µmol, 85%). ¹H NMR (D₂O, a series; protons of 5'-Gf, b series; protons of G-3') δ 7.94 (1H, s, H8), 7.85 (1H, s, H8), 6.10 (1H, dd, J = 3.0, 17.0 Hz, H1'a), 5.87 (1H, d, J = 5.5 Hz, H1'b), 5.48 (1H, ddd, J = 3.6, 4.1, 51.4 Hz, H2'a), 4.86–4.97 (1H, m, H3'a), 4.70 (1H, t, J = 5.2 Hz, H2'b), 4.49 (1H, dd, J = 4.7, 4.9 Hz, H3'b), 4.33 (1H, br s, H4'b), 4.28 (1H, br s, H4'a), 4.15–4.25 (2H, m, H5'b), 3.76–3.89 (2H, m, H5'a); UV λ_{max} (MeOH) nm: 252; FAB-MS m/z: 651 [M⁺-H].

 N^2 -Acethyl-2',3'-di-*O*-isobutyrylguanosine (7). This compound was prepared from 5'-*O*-trityl- N^2 -acetylguanosine by successive treatment with isobutyryl chloride and acid treatment in 76% yield. ¹H NMR (CDCl₃) δ 12.20 (1H, s, N^1 -H), 9.54 (1H, s, N^2 -H), 7.74 (1H, s, H8), 5.88–5.97 (2H, m, H1', H2'), 5.67 (1H, dd, J = 2.7, 5.2 Hz, H3'), 4.26 (1H, d, J = 2.2 Hz, H4'), 3.95 (1H, dd, J = 2.2, 12.6 Hz, H5'a), 3.82 (1H, dd, J = 1.6, 12.6 Hz, H5'b), 2.47–2.68 (2H, m, CH x2), 2.33 (3H, s, Ac), 1.08–1.23 (12H, m, CH₃ × 4); UV λ_{max} (MeOH) nm: 257, 280; EI-MS m/z: 465 [M⁺].

REFERENCES

- 1. Ledoux, L.; Baltus, E. Experientia **1954**, 10, 500–501.
- 2. Ledoux, L. Nature 1955, 175, 258-259.
- 3. Ledoux, L. Nature **1955**, *176*, 36–37.
- 4. a) Youle, R.J.; D'Alessio, G. *Ribonucleases: Structures and Funcions*; Academic Press: New York, 1997; 491–514. b) Irie, M.; Nitta, K.; Nonaka, T. Cell. Mol. Life Sci. **1998**, *54*, 775–784.
- 5. Vitagliano, L.; Adinolfi, S.; Riccio, A.; Sica, F.; Zagari, A. Protein Sci. **1998**, 7, 1691–1699.
- 6. Vitagliano, L.; Merlino, A.; Zagari, A.; Mazzarella, L. Protein Sci. **2000**, *9*, 1217–1225.
- 7. Aguilar, C.F.; Thomas, P.J.; Mills, A.; Moss, D.S.; Palmer, R.A. J. Mol. Biol. **1992**, *224*, 265–267.
- 8. Koepke, J.; Maslowska, M.; Heinemann, U.; Saenger, W. J. Mol. Biol. **1989**, *206*, 475–488.
- 9. Nonaka, T.; Nakamura, K.T.; Uesugi, S.; Ikehara, M.; Irie, M.; Mitsui, Y. Biochemistry **1993**, *32*, 11,825–11,837.
- 10. a) Ikehara, M.; Maruyama, T.; Miki, H. Tetrahedron **1978**, *34*, 1133–1188. b) Ikehara, M. Heterocycles **1984**, *21*, 75–90 and the references cited therein.

11. Enzymatic synthesis of purine 2'-deoxy-2'-fluororibosides has been reported by Welcome Research Laboratories. Tuttle, J.V.; Tisdale, M.; Krenitsky, T.A. J. Med. Chem. **1993**, *36*, 119–125.

- 12. Olsen, D.B.; Benseler, H.A.; Pieken, W.A.; Eckstein, F. Biochemistry **1991**, *30*, 9735–9741.
- 13. a) Fukukawa, K.; Ueda, T.; Hirano, T. Chem. Pharm. Bull. **1983**, *31*, 1582–1592. b) Sato, Y.; Maruyama, T.; Honjo, M. ibid. **1989**, *37*, 1604–1608.
- 14. Robins, M.J.; Wilson, J.S.; Sawyer, L.; James, M.N.G. Can. J. Chem. **1983**, *61*, 1911–1920.
- 15. Baker, D.C.; Kumar, S.D.; Waites, W.J.; Arnett, G.; Shannon, W.M.; Higuchi, W.I.; Lambert, W.J. J. Med. Chem. **1984**, *27*, 270.
- a) Ikehara M.; Maruyama, T. Tetrahedron 1975, 31, 1369–1372. b) Ikehara, M.;
 Miki, H. Chem. Pharm. Bull. 1978, 26, 2449–2453. c) Kawasaki, A.M.; Casper,
 M.D.; Freier, S.M.; Lesnik, E.A.; Zounes, M.C.; Cummins, L.L.; Gonzalez,
 C.; Cook, P.D. J. Med. Chem. 1993, 36, 831–841.
- 17. Atkinson, T.; Smith, M.A Practical Approach. In *Oligonucleotide synthesis*; Gait, M.J., Ed.; IRL Press: Oxford, 1984; 35–81.

Received February 20, 2002 Accepted April 15, 2002